

## Supplementary Information

### **A low-cost recombinant glycoconjugate vaccine confers immunogenicity and protection against enterotoxigenic *Escherichia coli* infections in mice**

Asher J. Williams<sup>1</sup>, Katherine F. Warfel<sup>2,3,4</sup>, Primit Desai<sup>5</sup>, Jie Li<sup>6</sup>, Jen-Jie Lee<sup>6</sup>, Derek A. Wong<sup>2,3,4</sup>, Phuong M. Nguyen<sup>1</sup>, Yufan Qin<sup>1</sup>, Sarah E. Sobol<sup>2,3,4</sup>, Michael C. Jewett<sup>2,3,4,7</sup>, Yung-Fu Chang<sup>6</sup> and Matthew P. DeLisa<sup>1,5,8\*</sup>

<sup>1</sup>Robert F. Smith School of Chemical and Biomolecular Engineering, Cornell University, Ithaca, NY 14853 USA

<sup>2</sup>Department of Chemical and Biological Engineering, Northwestern University, 2145 Sheridan Road, Technological Institute E136, Evanston, IL 60208, USA

<sup>3</sup>Chemistry of Life Processes Institute, Northwestern University, 2170 Campus Drive, Evanston, IL 60208, USA

<sup>4</sup>Center for Synthetic Biology, Northwestern University, 2145 Sheridan Road, Technological Institute E136 Evanston, IL 60208, USA

<sup>5</sup>Biochemistry, Molecular and Cell Biology, Cornell University, Ithaca, NY 14853

<sup>6</sup>Department of Population Medicine and Diagnostic Sciences, College of Veterinary Medicine, Cornell University, Ithaca, NY, 14853 USA

<sup>7</sup>Department of Bioengineering, Stanford University, Stanford, CA 40305 USA.

<sup>8</sup>Cornell Institute of Biotechnology, Cornell University, Ithaca, NY 14853 USA

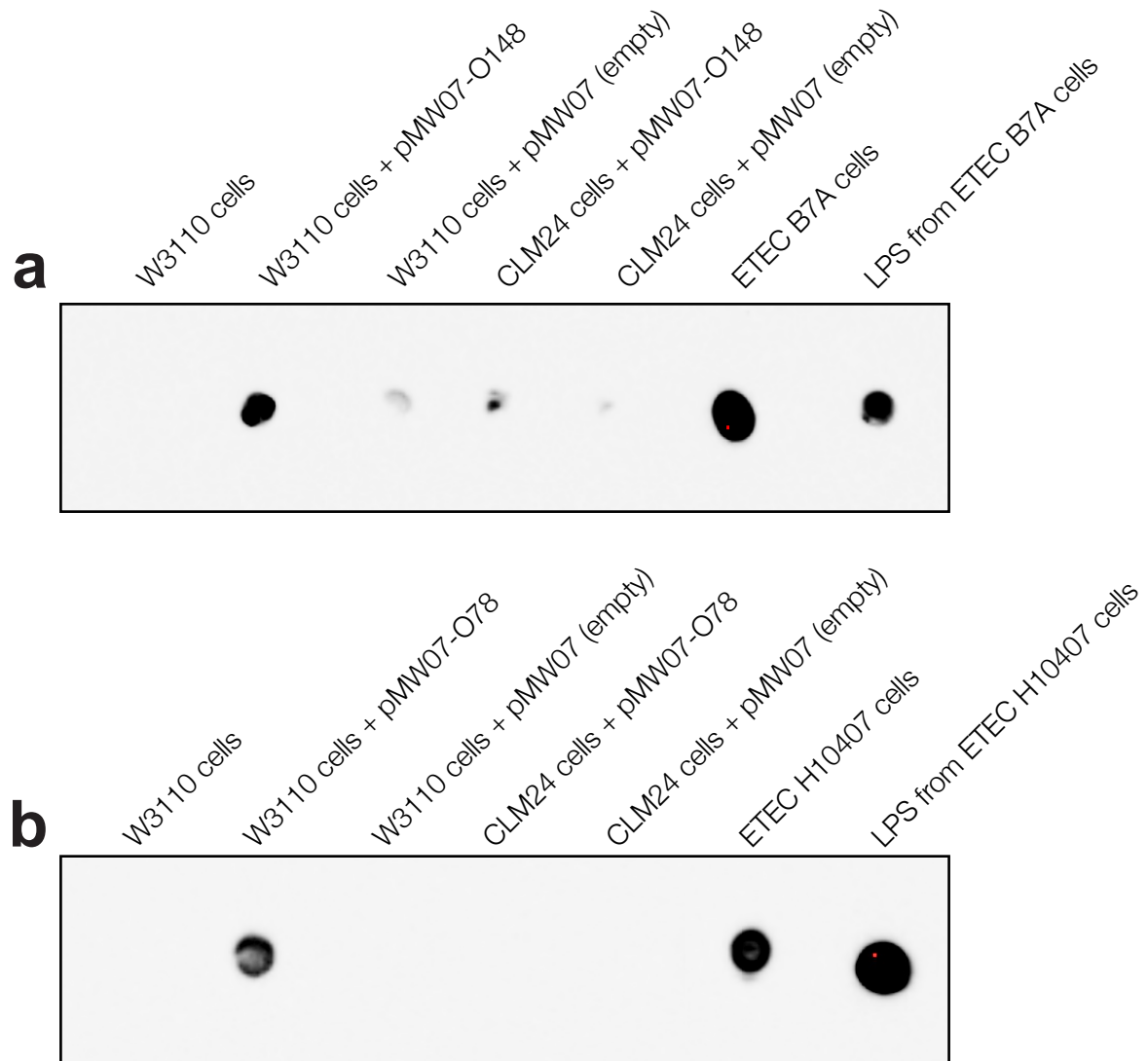
\*Address correspondence to: Matthew P. DeLisa, Robert Frederick Smith School of Chemical and Biomolecular Engineering, Cornell University, Ithaca, NY 14853 USA. Tel: 607-254-8560; Email: [md255@cornell.edu](mailto:md255@cornell.edu)

**Supplementary Table 1.** Strains and plasmids used in this study.

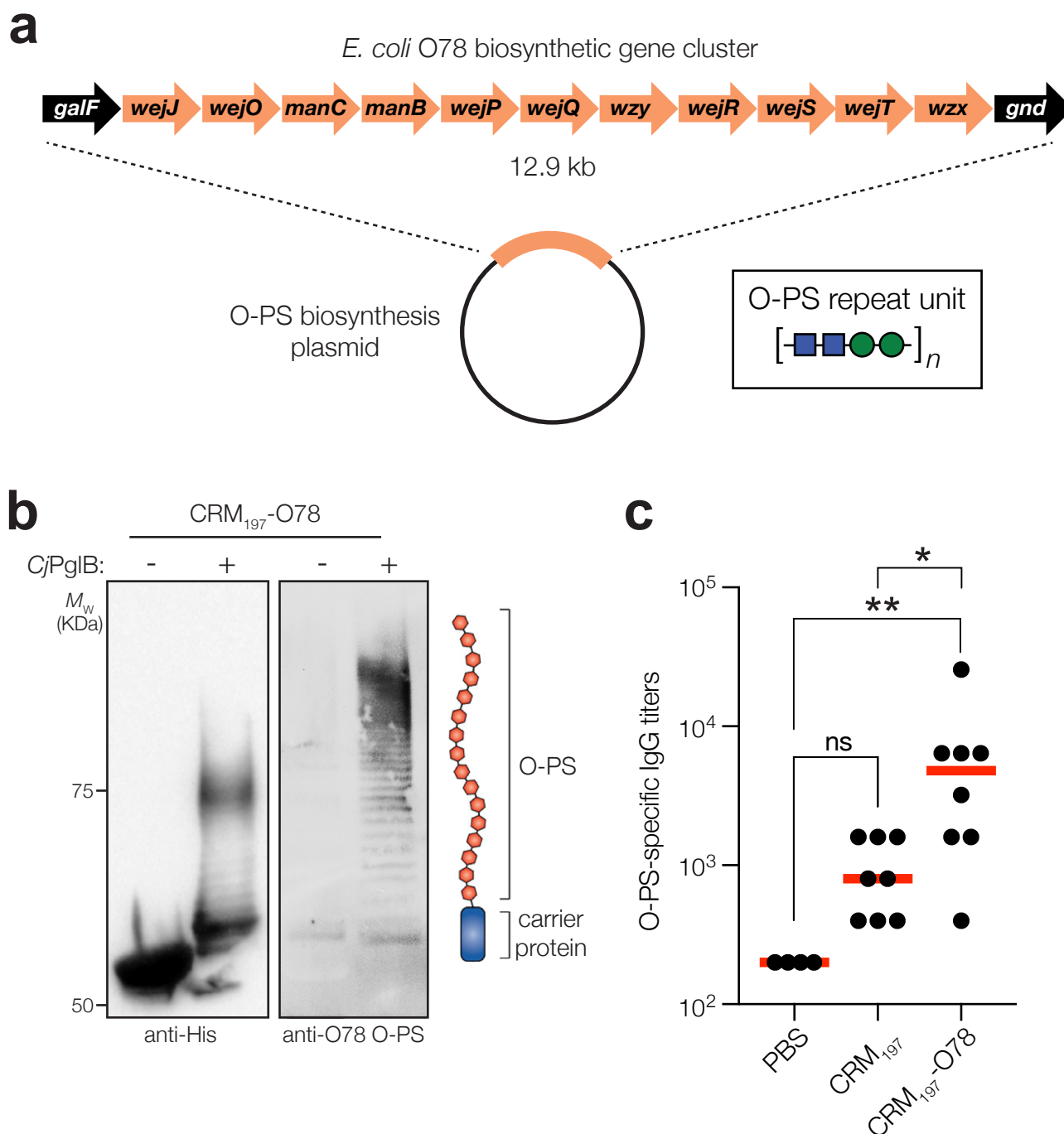
Strain or Plasmid	Description	Source or Reference
<i>E. coli</i> DH5α	<i>supE44 Δ(lacZYA-argF)U196 (Φ80ΔlacZM15) hsdR17 recA1 endA1 gyrA96 thi-1 relA1</i>	Lab stock
<i>E. coli</i> W3110	F <sup>-</sup> λ <sup>-</sup> <i>mcrA mcrB IN(rrnD-rnE)1</i>	Lab stock
<i>E. coli</i> CLM24	<i>E. coli</i> W3110 with deletion in the <i>waaL</i> gene encoding the O-antigen ligase	1
<i>E. coli</i> CLM24 Δ <i>lpxM</i>	<i>E. coli</i> CLM24 with knockout of the <i>lpxM</i> gene encoding the lipid A acyltransferase	2
ETEC strain B7A	O148:H28; CS6, LT, STa	3
ETEC strain H10407	O78:H11; CFA/I, LT, STa	4
pTrc99A-ssDsbA-PD <sup>4xDQNAT</sup>	<i>H. influenzae</i> protein D modified at N-terminus with signal peptide derived from <i>E. coli</i> DsbA and modified at C-terminus with 4x repeat of DQNAT glycosylation sequon and a 6x-His tag in the pTrc99A expression vector, Amp <sup>R</sup>	This study
pTrc99A-ssDsbA-CRM <sub>197</sub> <sup>4xDQNAT</sup>	Non-toxic mutant of <i>Corynebacterium diphtheriae</i> diphtheria toxin modified at N-terminus with signal peptide derived from <i>E. coli</i> DsbA and modified at C-terminus with 4x repeat of DQNAT glycosylation sequon and a 6x-His tag in the pTrc99A expression vector, Amp <sup>R</sup>	This study
pMAF10	HA-tagged CjPglB cloned in pMLBAD, Tmp <sup>R</sup>	1
pMAF10 <sup>D54N/E316Q</sup>	HA-tagged mutant CjPglB with D54N/E316Q mutations cloned in pMLBAD, Tmp <sup>R</sup>	5
pMW07-O148	<i>E. coli</i> O148 O-PS gene cluster in the pMW07 expression vector, Cm <sup>R</sup>	6, 7
pMW07-O78	<i>E. coli</i> O78 O-PS gene cluster in the pMW07 expression vector, Cm <sup>R</sup>	2, 7, 8
pJL1-PD <sup>4xDQNAT</sup>	PD carrier protein modified at C-terminus with 4x repeat of DQNAT glycosylation sequon and a 6x-His tag in the pJL1 expression vector, Kan <sup>R</sup>	2
pSF-CjPglB-LpxE	CjPglB along with <i>Francisella tularensis</i> LpxE that promotes monophosphorylation of lipid A, Amp <sup>R</sup>	27

## References

1. Feldman, M.F. et al. Engineering N-linked protein glycosylation with diverse O antigen lipopolysaccharide structures in *Escherichia coli*. *Proc Natl Acad Sci U S A* **102**, 3016-3021 (2005).
2. Stark, J.C. et al. On-demand biomanufacturing of protective conjugate vaccines. *Sci Adv* **7** (2021).
3. DuPont, H.L. et al. Pathogenesis of *Escherichia coli* diarrhea. *N Engl J Med* **285**, 1-9 (1971).
4. Levine, M.M. et al. Lack of person-to-person transmission of enterotoxigenic *Escherichia coli* despite close contact. *Am J Epidemiol* **111**, 347-355 (1980).
5. Ollis, A.A., Zhang, S., Fisher, A.C. & DeLisa, M.P. Engineered oligosaccharyltransferases with greatly relaxed acceptor-site specificity. *Nat Chem Biol* **10**, 816-822 (2014).
6. Celik, E. et al. Glycoarrays with engineered phages displaying structurally diverse oligosaccharides enable high-throughput detection of glycan-protein interactions. *Biotechnol J* **10**, 199-209 (2015).
7. Chen, L. et al. Outer membrane vesicles displaying engineered glycotopes elicit protective antibodies. *Proc Natl Acad Sci U S A* **113**, E3609-3618 (2016).
8. Warfel, K.F. et al. A low-cost, thermostable, cell-free protein synthesis platform for on demand production of conjugate vaccines. *bioRxiv*, 2022.2008.2010.503507 (2022).



**Supplementary Figure 1. Biosynthesis of ETEC O-PS antigens in non-pathogenic *E. coli*.** Dot blot analysis of whole cells and LPS derived from the strains indicated. A total of 2  $\mu$ L of OD<sub>600</sub>-matched cultures or LPS extracts were spotted on nitrocellulose membranes. Membranes were probed with (a) anti-ETEC O148 antibody or (b) anti-ETEC O78 antibody. Results are representative of three biological replicates.



**Supplementary Figure 2. Biosynthesis of ETEC O78 O-PS antigen in non-pathogenic *E. coli*.** (a) Biosynthesis of ETEC O78 O-PS from plasmid pMW07-O78, which encodes the entire O-PS locus from ETEC strain H10407 (serotype O78:H11) between *galF* and *gnd*. (b) Immunoblot analysis of purified carrier proteins derived from *E. coli* CLM24  $\Delta p x M$  cells carrying a plasmid encoding CRM<sub>197</sub><sup>4xDQ<sub>NAT</sub></sup> along with plasmid pMW07-O78 encoding the ETEC O78 O-PS biosynthetic pathway and plasmid pMAF10 encoding wild-type CjPglB (wt) or an inactive mutant of CjPglB (mut) as indicated. Blots were probed with anti-His antibody to detect acceptor proteins and anti-ETEC O78 antibody to detect O-PS. Images depict aglycosylated and multiply glycosylated forms of CRM<sub>197</sub><sup>4xDQ<sub>NAT</sub></sup>. Molecular weight ( $M_w$ ) markers are indicated on the left. Results are representative of three biological replicates. (c) O-PS-specific IgG titers in day 56 serum of individual mice (black circles) and mean titers of each group (red lines) as determined by ELISA with LPS derived from ETEC H10407 as immobilized antigen. Groups of three BALB/c mice were immunized s.c. with 100  $\mu$ L PBS alone or PBS containing 50  $\mu$ g of either aglycosylated CRM<sub>197</sub><sup>4xDQ<sub>NAT</sub></sup> carrier protein or glycosylated CRM<sub>197</sub><sup>4xDQ<sub>NAT</sub></sup> bearing ETEC O78 O-PS adjuvanted with aluminium phosphate adjuvant. Mice were boosted on days 21 and 42 with identical doses of each immunogen. Data were analyzed for statistical significance using the Mann-Whitney test (\* $p < 0.05$ ; \*\* $p < 0.01$ ; ns, not significant).